

# Immunofluorescent Staining of Mouse and Rat Leukocytes

## I. Procedure

1. Harvest cells from tissue, preparing a single cell suspension. Red blood cells may be removed by lysis or density gradient: Red blood cells from murine peripheral blood or a spleen cell suspension can be lysed using BD Biosciences Pharmingen's PharM Lyse™ (Cat. No. 555899) solution. Add 2.0 ml of 1X Lysing Solution to the spleen cell suspension or per 200  $\mu$ l of murine peripheral blood. Gently vortex immediately after adding the lysing solution. Incubate at room temperature, protected from light, for 3-4 min for mouse cells or 5-6 min for rat cells. Centrifuge 200 x g for 5 min. Carefully aspirate and dispose of supernatant, without disturbing the pellet. Resuspend pellet in 1X cold wash buffer (PBS/0.1% NaN<sub>3</sub>/1.0% fetal bovine serum). Centrifuge at 350 x g for 5 min. Finally, resuspend cell pellet to a concentration of 2 x 10<sup>7</sup> cells/ml (i.e., 10<sup>6</sup> cells per 50  $\mu$ l).
2. Dilute primary mAbs (e.g., unconjugated, biotinylated, or fluorochrome-conjugated mAbs) to predetermined optimal concentrations in wash buffer and deliver to the wells of a U-bottom microtiter plate in a volume of 50  $\mu$ l.
3. Deliver 10<sup>6</sup> cells in 50  $\mu$ l to each well already containing 50  $\mu$ l of mAb (or 50  $\mu$ l wash buffer for negative controls). Mix by gently vortexing or tapping.
4. Incubate at 4°C for 20-40 min in the dark.
5. Wash 2X with 200  $\mu$ l wash buffer (or 3X if a biotin-conjugated primary antibody is used). After each centrifugation, 350 x g for 5 min, aspirate wells or flick plate to remove supernatant. Vortex gently or tap plate to loosen pellet prior to adding next wash or diluted secondary reagent.
6. If a second-step reagent is needed, resuspend cell pellet in 100  $\mu$ l of appropriately diluted secondary reagent (e.g., fluorochrome-conjugated avidin, streptavidin, anti-Ig allotype, anti-Ig isotype, polyclonal anti-Ig). For example, dilute antibody to ~1  $\mu$ g per 100  $\mu$ l in wash buffer and add this to each well containing the loosened cell pellet.
7. Incubate at 4°C for 20-40 min in the dark.
8. Wash 2X with 200  $\mu$ l wash buffer, as in Step 5. Use 100  $\mu$ l wash buffer to transfer cell pellets to 0.4 ml aliquots of wash buffer (final concentration ~10<sup>6</sup> cells in 0.5 ml) in tubes appropriate for flow cytometer. Acquire sample data on flow cytometer as soon as possible after staining.